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Mukherjee, Nobina; Kocer, Armagan

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- [1] Perozo E et al. (2002). Open channel structure of MscL and the gating mechanism of mechanosensitive channels. *Nature*, 418, 942-948
- [2] Corry B et al. (2010). An improved open-channel structure of MscL determined from FRET confocal microscopy and simulation. *Journal of General Physiology*, 136, 483-494

#### 613-Pos Board B399

##### Activation of Ms Channel by LPC: A Trigger for MscL Gating in the Absence of Applied Tension

Nobina Mukherjee, Armagan Kocer.

Groningen Biomolecular Sciences and Biotechnology Institute, University of Groningen, Groningen, Netherlands.

Mechanosensitive Channel of Large Conductance (MscL) allows bacteria to respond to osmotic stress in the environment. It senses the increase in the lateral pressure in the membrane due to sudden hypo-osmotic shock and acts as a safety valve. MscL has one of the largest pores in nature; in its open state it allows the passage of ions and small molecules upto 6.5 kDa. MscL has been used in this study as an externally controlled valve i.e. the opening of the channel is controlled by external stimuli.

Several techniques like patch clamp, EPR spectroscopy has been applied towards elucidating the gating mechanism of MscL. EPR is effective in tracking the initial conformational changes that the protein may undergo during gating. The main challenge in using spectroscopy is that, unlike patch clamp technique, tension cannot be applied directly for opening the channel. L- $\alpha$ -lysophosphatidylcholine, a reported activator of MscL was studied in this work to trigger opening of the channel in a controlled way. In our work we provide evidence that LPC mimics tension in opening the channel. Our findings also clearly show that LPC can be used for phenotypic characterization of MscL mutants, in a much simpler experiment than patch clamp. A clear differentiation in activity between GOF, LOF and Wt *Ec* MscL is observed at 4  $\mu$ M LPC. In conclusion, we characterized an activator with which the mechanism of channel gating can be studied in a controlled way.

#### 614-Pos Board B400

##### Piezo1 Gating: Comparison Between Whole Cell Currents and the Patch

Philip Gottlieb, Chilman Bae, Frederick Sachs.

SUNY at Buffalo, Buffalo, NY, USA.

Piezo1 channels gate with mechanical stress in the membrane and gating involves both activation and inactivation. In HEK293 cells transfected with Piezo1 and subjected to pressure stimuli, cell-attached patch recordings showed that the inactivation rate slowed as extracellular divalent ions were reduced. With  $>1\text{mM Mg}^{2+}$ , activation had no measurable latency and the inactivation rate was rapid but stress dependent, suggesting that  $\text{Mg}^{2+}$  may act as an open channel blocker (the effects of  $\text{Ca}^{2+}$  are in progress). Without divalents there was no inactivation, but surprisingly, activation now had a pronounced latency ( $\sim 500$  ms). Inactivation may actually represent adaptation of the local stimulus by the cytoskeleton and not overt channel closure. To disrupt the cytoskeleton we treated cells with cytochalasin D before patching and found inactivation was unaffected suggesting cytoskeletal adaption was not the cause. Attempting the inverse experiment, we increased cytoskeletal stress by swelling the cells osmotically, but that too didn't affect the inactivation rate.

For analogy close to the in situ situation, we evoked whole cell Piezo1 currents by indenting cells with a glass probe. Like the patch, removing extracellular divalent ions reversibly reduced the inactivation rate. However, in contrast to patch recordings, Cytochalasin D caused a loss of whole cell current and cell swelling increased the evoked currents. These results suggest that the forces that gate Piezo1 in whole cell mode propagate through the cytoskeleton, and that divalent ion block may be responsible for inactivation.

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#### 615-Pos Board B401

##### A High-Throughput Technique for Screening Novel Antibacterial Agents Targeting Bacterial Mechanosensitive Channels

Bruce A. Cornell<sup>1</sup>, Andrew R. Battle<sup>2,3</sup>, Sonia Carne<sup>1</sup>, Boris Martinac<sup>3,4</sup>.

<sup>1</sup>SDxTethered Membranes PTL, St Leonards 2065, Australia, <sup>2</sup>Griffith University, Gold Coast Campus 4222, Australia, <sup>3</sup>Victor Chang Cardiac Research Institute, Darlinghurst 2010, Australia, <sup>4</sup>St Vincents Clinical School, The University of New South Wales, Sydney, Australia.

Mechanosensitive Ion channels are essential for maintaining cellular homeostasis. It has been proposed that they function as osmotically activated emergency valves, guarding against membrane rupture by opening pores to release excessive turgor pressure. Interfering with their function suggests many novel mechanisms to complement conventional antimicrobials to kill or inhibit growth of bacterial pathogens. Typical of this class of mechanosensitive channel are the large conduction homopentameric bacterial channel, MscL (MW $\sim$ 17kDa) and the small conduction homoheptameric bacterial channel, MscS ( $\sim$ 37kDa).

These respond respectively to large and small osmotic pressures, and elicit ion conductances of  $\sim 3\text{nS}$  and  $\sim 1\text{nS}$ . Both MscL and MscS exhibit strong homology across all bacteria. In this study we report the use of a.c. impedance spectroscopy of pure MscL and MscS channels in a family of tethered bilayer membrane systems as a high-throughput technique that can be used to screen for potential lead compounds for the development of novel antibacterial agents that interfere with the function of MS channels. Using either a swept frequency Bode profile or a single frequency impedance measure, plates of 96 electrodes may be screened simultaneously. Typical conditions are a measurement of resistance in the range of 1k to 100M in response to excitation over frequency in the range of 0.1Hz to 1 kHz. The robustness of the tethered membrane permits modulation of the MS channel conductance through an alteration in the membrane thickness. This can be achieved through the application of large transmembrane potentials or the dilution of the membrane lipids with surfactants possessing varying hydrophobic chain lengths.

#### 616-Pos Board B402

##### Mechanotransduction in A549 Alveolar Cells via Cell Stretch-Induced ATP Release

Ryszard Grygorczyk<sup>1</sup>, Kishio Furuya<sup>2</sup>, Masahiro Sokabe<sup>3</sup>.

<sup>1</sup>University of Montreal, Montreal, QC, Canada, <sup>2</sup>Nagoya University, Nagoya, Japan, <sup>3</sup>Nagoya University Graduate School of Medicine, Nagoya, Japan.

Mechanotransduction at the cellular/tissue level often involves release of signaling molecules. Among them, purines appear to be the most primitive and widespread chemical messengers in the animal and plant kingdoms. Their release is highly mechano-sensitive, but the release pathways and regulatory mechanisms are not well understood. Here, we investigated the effect of unidirectional stretch on ATP release from human lung A549 alveolar cells grown on a flexible substrate. We used real-time luciferin-luciferase bioluminescence imaging combined with IR imaging to simultaneously monitor cellular ATP release and extend of cell stretch. Single 1-s stretch of 15-30% induced transient ATP release that ceased in 2-3 min and was restricted to a limited number of cells. The number of responding cells increased dose-dependently with the extent of stretch but did not involve cell damage. Calibration of the ATP response showed that local ATP concentration in the close proximity ( $\leq 150\text{ }\mu\text{m}$ ) to stretch-activated cells may exceed 1  $\mu\text{M}$  or even 10  $\mu\text{M}$ . These concentrations are sufficient for autocrine/paracrine stimulation of cell surface purinergic receptors on the neighboring cells. ATP responses were insensitive to putative ATP channel blockers carbenoxolone or NPPB (100  $\mu\text{M}$ ), inhibitors of pannexin or anion channels respectively, but were abolished by N-ethylmaleimide. Fluo8 fluorescence measurement of stretch-induced intracellular  $\text{Ca}^{2+}$  responses revealed that limited number of cells displayed rapid responses, which peaked in  $<1\text{-s}$  and ceased in 1-3 min. This is similar to stretch-induced ATP responses and suggests functional connection between the two signals. Experiments show that cell stretch induces ATP release via cell-regulated process, likely exocytosis. Mechano-sensitive ATP release, via autocrine/paracrine effects, initiates purinergic signaling cascade in other cells and may function as a general intercellular mechanotransduction paradigm in the lung and other tissues.

#### 617-Pos Board B403

##### Coarse-Grained Molecular Dynamics Simulation Study Focusing on the Conformational Changes of Transmembrane Helices of the E-Coli Mechanosensitive Channel MscL

Hidetoshi Matsuo<sup>1</sup>, Yasuyuki Sawada<sup>1</sup>, Masahiro Sokabe<sup>1,2</sup>.

<sup>1</sup>Department of Physiology, Nagoya University Graduate School of Medicine, Nagoya, Japan, <sup>2</sup>First Research Center for Innovative Nanobiodevice, Nagoya University, Nagoya, Japan.

Mechanosensitive channels are activated by sensing membrane tension. MscL is a homopentamer of a subunit with transmembrane inner (TM1) and outer (TM2) helices and TM1s line the ion/water permeable pore. We have analyzed the gating properties of MscL using patch-clamp experiments and simulations such as all-atom (AA) molecular dynamics (MD). However, in AA MD simulations, we need to apply about 10 times bigger membrane tension than experimentally applied to open the pore. In this study, we performed coarse-grained (CG) MD simulations to reproduce the opening process of MscL under appropriate conditions based on the experimental ones and to find the differences of the conformational changes of transmembrane helices between wild type (WT) and mutant models (F78N and G22N). We constructed CG MscL model based on the equilibrated all-atom closed model. After constructing, MscL was embedded in the lipid bilayer and water beads were added. Then we performed CG MD simulations for the opening of MscL under 60 bar, about three times bigger membrane tension for 1  $\mu\text{s}$ . Upon membrane stretch, all types of MscL could open its pore during the simulations. In order to analyze the correlation of the conformational changes between TM1 and TM2 helices, we